amber with four black fascia on disc and another pair near middle of sides almost attaining the lateral margins, the anterior discal fascia do not attain the anterior margin and are bluntly oval, the posterior pair are convergent posteriorly and are produced to attain the posterior margin. Elytra amber colored with black areas disposed as follows: humeral angles and scutellum and its cavity forming an irregular transverse band across the base; elongate area on each side margin near middle with a sinuate band connecting them with the median vitta which extends backward from the scutellum; posterior third except for two lateral rounded spots. Prosternum and flanks amber, prosternal sutures black, remainder of ventron dark brown, legs except coxæ yellow. Front convex, anterior margin broadly rounded, moderately punctate. Antennæ with joints two and three subequal, together equal to four. Pronotum not tuberculate at base, moderately evenly sparsely punctate; posterior angles unicarinate, carina distinct to apex of posterior angles where it joins the lateral margin. Elytra not spinose at tip. Prosternum finely punctate, flanks coarsely punctate. Last abdominal segment similar in punctuation to preceding segments. Tarsal joint four (Pl. V, Fig. 3b) with a moderately long slender lobe, not visible from above.

Length, 5 mm.

Type locality: Rio Madeira, Brazil.

Type: No. 9046 Mus. Comp. Zoöl. Harvard University.

Type collected by W. M. Mann and F. Baker.

This species which I have named in honor of Dr. Baker is near *Monocrepidius pictus* Cand. but is easily distinguished from this species by the absence of the basal pronotal tubercle.

EXPLANATION OF PLATE V.

Fig. 1. *Drasterius manni;* a. dorsum of adult; b. pronotal sculpturing; c. posterior coxal plate; d. propleural sculpturing; e. tarsus; f. tarsal claws.

Fig. 2. *Monocrepidius madeirensis;* a. dorsum of adult; b. second, third, and fourth antennal joints; c. third, fourth, and fifth tarsal joints.

Fig. 3. *Monocrepidius bakeri;* a. dorsum of adult; b. tarsus.

A PSYLLID GALL ON JUNCUS (*LIVIA MACULIPENNIS* FITCH).[1]

By Edith M. Patch.

Since 1857 when Fitch described this beautiful little insect, nothing more definite in regard to its habits has been recorded than that it is found in swampy places. Thomas in 1879 said that it was "found on the sweet-flag," but there is nothing in his account which would indicate that it fed upon that plant.

[1] Papers from the Maine Agricultural Experiment Station: Entomology, No. 82.

This past summer, however, the secret of its habitation was discovered by Miss Cora H. Clarke of Boston, who made an interesting collection of Juncus galls near Magnolia Village, Mass., on August 17, which she shared with the writer. At this date the galls contained only unknown nymphs of a Psyllid but they were about ready to wing and the adults began to emerge in large numbers on August 20–21. These proved to be *Livia maculipennis* Fitch.

The accompanying photograph gives the enlarged abnormal growth of the galled-specimen in contrast to the normal rush, and makes a description unnecessary.

Did Fitch, nearly sixty years ago, pick a stem of rush with its monstrous tassel deserted by its colony and wonder "What did that?" while the little "spotted wings" were flitting about the marsh or resting perchance upon sweet-flag near by?

THE COCCIDÆ OF NEW JERSEY GREENHOUSES.

By Harry B. Weiss,
New Brunswick, New Jersey.

While greenhouse coccids are not strictly a part of the New Jersey fauna, yet many of them are more or less permanently established and are a constant source of annoyance and expense to various growers. Except for the "mealy bugs" and a few species of "soft" and "armored scales," many of them are difficult to control and most of the insecticides recommended for greenhouse use are ineffective. Many of them will kill larvæ but here their usefulness ends. Many have been recommended by hearsay, on the basis of too little investigation, or after experimentation of a meager kind which neglected to include atmospheric conditions and other factors of a more or less variable nature which have an important bearing on the success or failure of the material.

As a result, the most effective work in combating scale insects in greenhouses is done when the plants are overhauled and repotted. At this time badly infested plants are destroyed, infested leaves pulled off, and men and girls are placed at work scrubbing the leaves with tooth brushes, erasers and soft cloths or using pointed sticks to dislodge the scales in cracks and crevices. In other words,